



# Microbial diversity in a saline siliciclastic aquifer at the ATES exploration site Berlin-Adlershof

Julia Mitzscherling<sup>1</sup>, Lioba Virchow<sup>2</sup>, Martin Gitter<sup>3</sup>, Armando Alibrandi<sup>1</sup>, Simona Regenspurg<sup>2</sup>, Stefan Kranz<sup>2</sup> and Dirk Wagner<sup>1,4</sup>

<sup>1</sup>GFZ German Research Centre for Geosciences, Section Geomicrobiology, Potsdam, Germany; <sup>2</sup>GFZ German Research Centre for Geosciences, Section Geoenergy, Potsdam, Germany; <sup>3</sup>Technische Universität Berlin, Department of Applied Geochemistry, Berlin, Germany; <sup>4</sup>University of Potsdam, Institute for Geosciences, Potsdam, Germany

### **Motivation**

- Aquifer Thermal Energy Storages (ATES) can store excess energy (heat) and **provide** it when required
- microbial activity can impair the efficiency and integrity of ATES facilities in different ways
- rucial to identify microbial key players and monitor their activity

#### **Objectives**

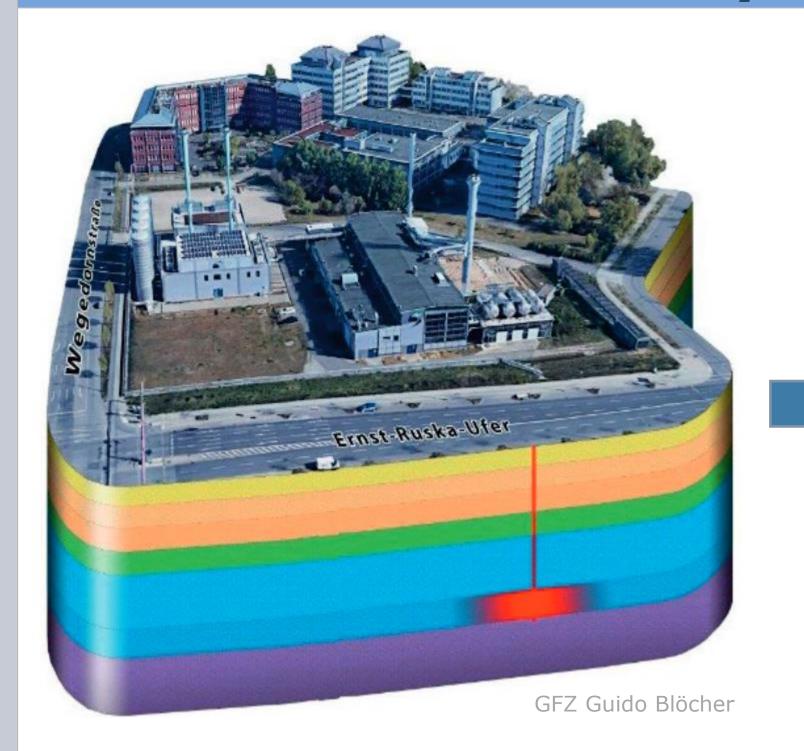
 explore microbial community composition, behaviour and potential functions in a siliciclastic aquifer at the ATES exploration site over 2 years after drilling

# Quarz Quarz

#### Take home

- Aquifer microbial community is characterized by syntrophic relationships between fermentative and acetogenic bacteria with sulfate reducing bacteria (SRB) and methanogens
- Community undergoes **succession over time** fermentation <del>></del> sulfate reduction → methanogenesis
- Temperature increase selects for SRB, decrease for acetogens
- Potential key consequences on ATES:
  - H<sub>2</sub>S formation → **corrosion** of metal components
  - Iron sulfide scales → pipe clogging
  - CH<sub>4</sub> formation → gas buildup/pressure changes/safety hazard
  - Biofilm formation -> permeability/effiency reduction

# **Study Site & Methodology**

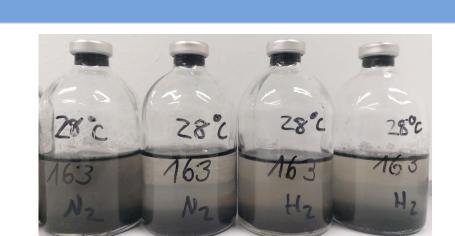


**ATES** exploration site\* in Berlin-Adlershof (BTB GmbH, Berlin)



- **Water sampling**
- 2 months
- 14 months 24 months
- 28 months after drilling

Origin of groundwater at ~225 mbs



**Enrichment for sulfate** reducers (SRB): +/- NaCl,  $N_2$  or  $H_2$  at



**Microbial** community analysis based on amplicon sequencing of 16S rRNA gene

28 months

14 months

stress: 0.10

24 months 🛧 💊

16/28/55 °C

# Hydrochemistry

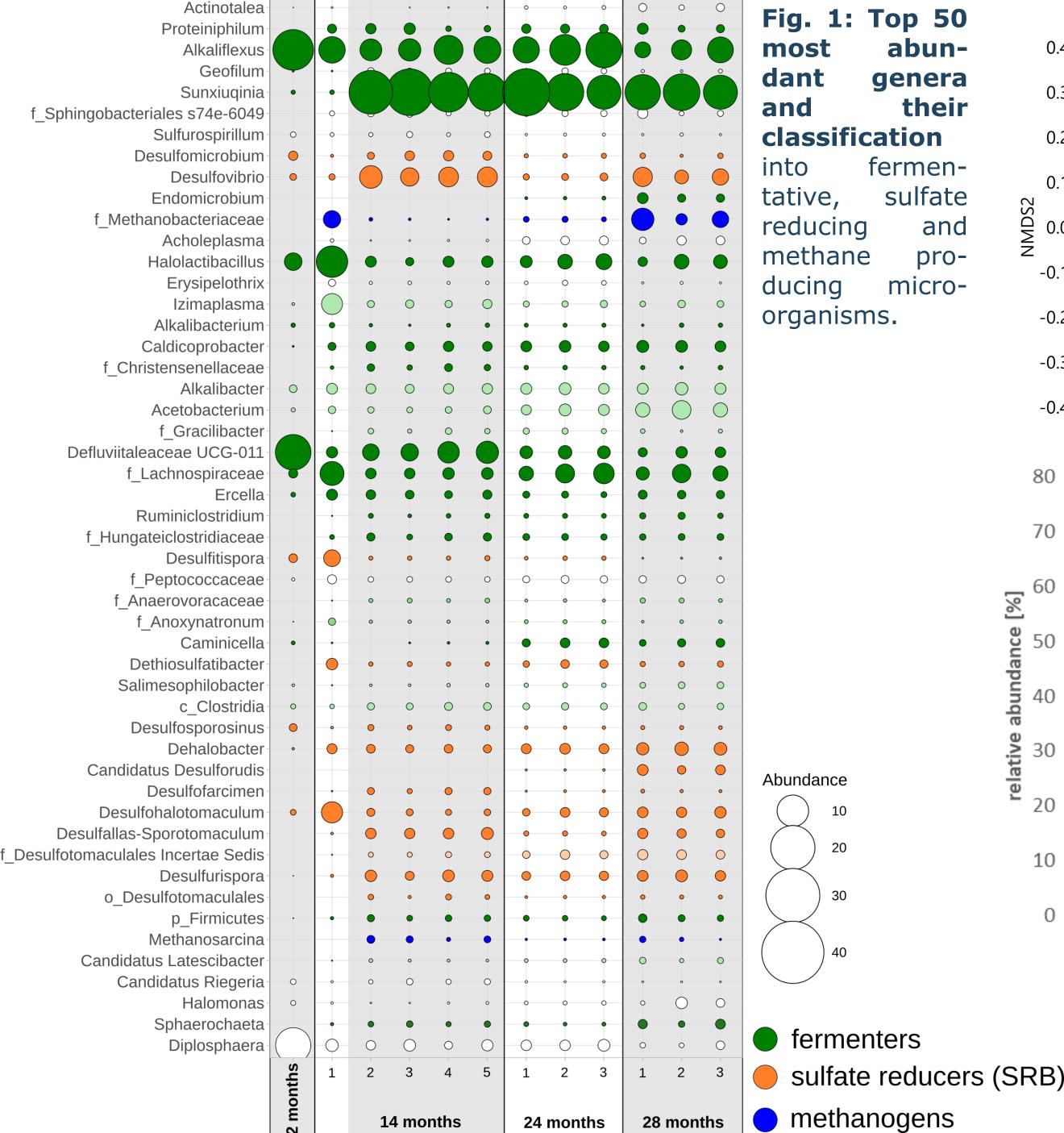
**Table 1: Groundwater characteristics.** Saline water  $(\sim 2\%)$  is dominated by Na and Cl. Sulfate can act as electron acceptor for anaerobic respiration by SRB. The presence of organic acids indicates fermentative microbial metabolism. \*Standing water in the annulus of the production well as indicated by higher pH, lower temperature and different microbial community (see Fig. 1 & 2A).

			LC		Lilig										
	sample		[mS	Temp	L-1]							Lactic			
sampling	no.	рН	cm <sup>-1</sup> ]	[° C]	Na	K	Mg	Ca	F	Cl	SO <sub>4</sub>	acid	Acetate	<b>Propionate</b>	<b>Valerate</b>
2 months	1	n.m.	31.9	15.9	6970	220	146	166	1	11688	270	n.a.	0.4	n.a.	n.a.
14 months	1*	9.5	26.4	11.8	6615	259	109	71	1	11163	270	<1	1.8	n.a.	<1
	2-5	7.7	31.9	13.6	6656	283	130	139	1	11060	250	3.6	4.0	0.6	0.9
24 months	1-3	7.6	31.4	14.1	5401	110	60	54	1	10457	235	0.3	< 0.1	< 0.1	n.a.
28 months	1-3	7.7	31.7	15.3	6843	152	151	190	n.a.	10770	225	<0.1	0.8	0.1	n.a.
	sampling 2 months 14 months 24 months	sampling no.  2 months 1  14 months 1*  2-5  24 months 1-3	2 months       1       n.m.         14 months       1*       9.5         2-5       7.7         24 months       1-3       7.6	sample sampling no.       [mS]         pH cm-1]       pH cm-1]         2 months       1       n.m. 31.9         14 months       1*       9.5       26.4         2-5       7.7       31.9         24 months       1-3       7.6       31.4	sample sampling no.       [mS Temp pH cm <sup>-1</sup> ][° C]         2 months       1         14 months       1*         2-5       7.7         31.9       13.6         24 months       1-3	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ] Na           2 months         1         n.m. 31.9 15.9 6970           14 months         1*         9.5 26.4 11.8 6615           2-5         7.7 31.9 13.6 6656           24 months         1-3         7.6 31.4 14.1 5401	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ] Na K           2 months         1         n.m. 31.9 15.9 6970 220           14 months         1*         9.5 26.4 11.8 6615 259           2-5         7.7 31.9 13.6 6656 283           24 months         1-3         7.6 31.4 14.1 5401 110	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ]         Na K Mg           2 months         1         n.m. 31.9 15.9 6970 220 146           14 months         1*         9.5 26.4 11.8 6615 259 109           2-5         7.7 31.9 13.6 6656 283 130           24 months         1-3         7.6 31.4 14.1 5401 110 60	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ]         Na K Mg Ca           2 months         1         n.m. 31.9 15.9 6970 220 146 166           14 months         1*         9.5 26.4 11.8 6615 259 109 71           2-5         7.7 31.9 13.6 6656 283 130 139           24 months         1-3         7.6 31.4 14.1 5401 110 60 54	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ]         Na K Mg Ca F           2 months         1         n.m. 31.9 15.9 6970 220 146 166 1           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1           2-5         7.7 31.9 13.6 6656 283 130 139 1           24 months         1-3         7.6 31.4 14.1 5401 110 60 54 1	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ]         Na K Mg Ca F Cl           2 months         1         n.m. 31.9 15.9 6970 220 146 166 1 11688           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1 11163           2-5         7.7 31.9 13.6 6656 283 130 139 1 11060           24 months         1-3         7.6 31.4 14.1 5401 110 60 54 1 10457	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         Na K Mg Ca F Cl SO <sub>4</sub> 2 months         1         n.m. 31.9 15.9 6970 220 146 166 1 11688 270           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1 11163 270           2-5         7.7 31.9 13.6 6656 283 130 139 1 11060 250           24 months         1-3         7.6 31.4 14.1 5401 110 60 54 1 10457 235	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L-1]         Lactic solution           2 months         1         n.m. 31.9 15.9 6970 220 146 166 1 11688 270 n.a.           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1 11163 270 <1	sample sampling no.         [mS Temp pH cm⁻¹][° C]         L⁻¹]         Na K Mg Ca F Cl SO₄ acid Acetate         Lactic acid Acetate           2 months         1         n.m. 31.9 15.9 6970 220 146 166 1 11688 270 n.a. 0.4           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1 11163 270 <1 1.8           2-5         7.7 31.9 13.6 6656 283 130 139 1 11060 250 3.6 4.0           24 months         1-3 7.6 31.4 14.1 5401 110 60 54 1 10457 235 0.3 <0.1	sample sampling no.         [mS Temp pH cm <sup>-1</sup> ][° C]         L <sup>-1</sup> ]         Lactic acid Acetate Propionate           2 months         1         n.m. 31.9 15.9 6970 220 146 166 1 11688 270 n.a.         n.a. 0.4 n.a.           14 months         1*         9.5 26.4 11.8 6615 259 109 71 1 11163 270 <1 1.8 n.a.           2-5         7.7 31.9 13.6 6656 283 130 139 1 11060 250 3.6 4.0 0.6           24 months         1-3 7.6 31.4 14.1 5401 110 60 54 1 10457 235 0.3 <0.1 <0.1

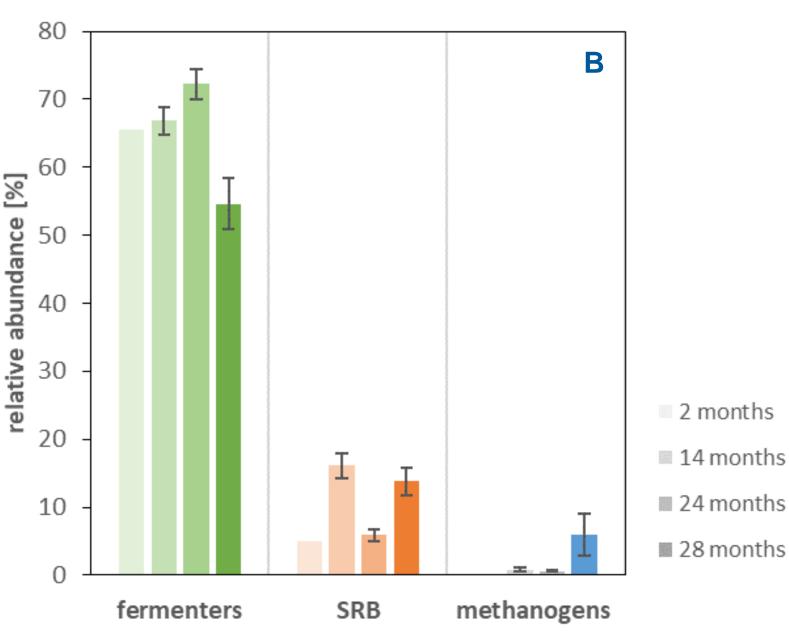
**Table 2: Dissolved gas composition –** dominated by H<sub>2</sub> 2 months after drilling, but N<sub>2</sub> 14 months after drilling.

	[%] H <sub>2</sub>	$N_2$	CO <sub>2</sub>	Ar	02	He	CH <sub>4</sub>
2 months	91.6	7.0	1.3	n.a.	n.a.	n.a.	n.a.
14 months	0.1	95.8	0.5	2.6	0.7	0.0	0.2

# **Aquifer Microbial Community Composition and Dynamics**



0.4 -14 months-annulus 0.3



-0.2

NDMS1

Fig. 2A: NMDS analysis microbial **community** shift after > 2 months and stabilization > 14 months. Relative abundance changes fermenters, SRB and methanogens over

https://www.gfz-potsdam.de/sektion/geoenergie/projekte/2019-2022-geofern

# **Enrichments**

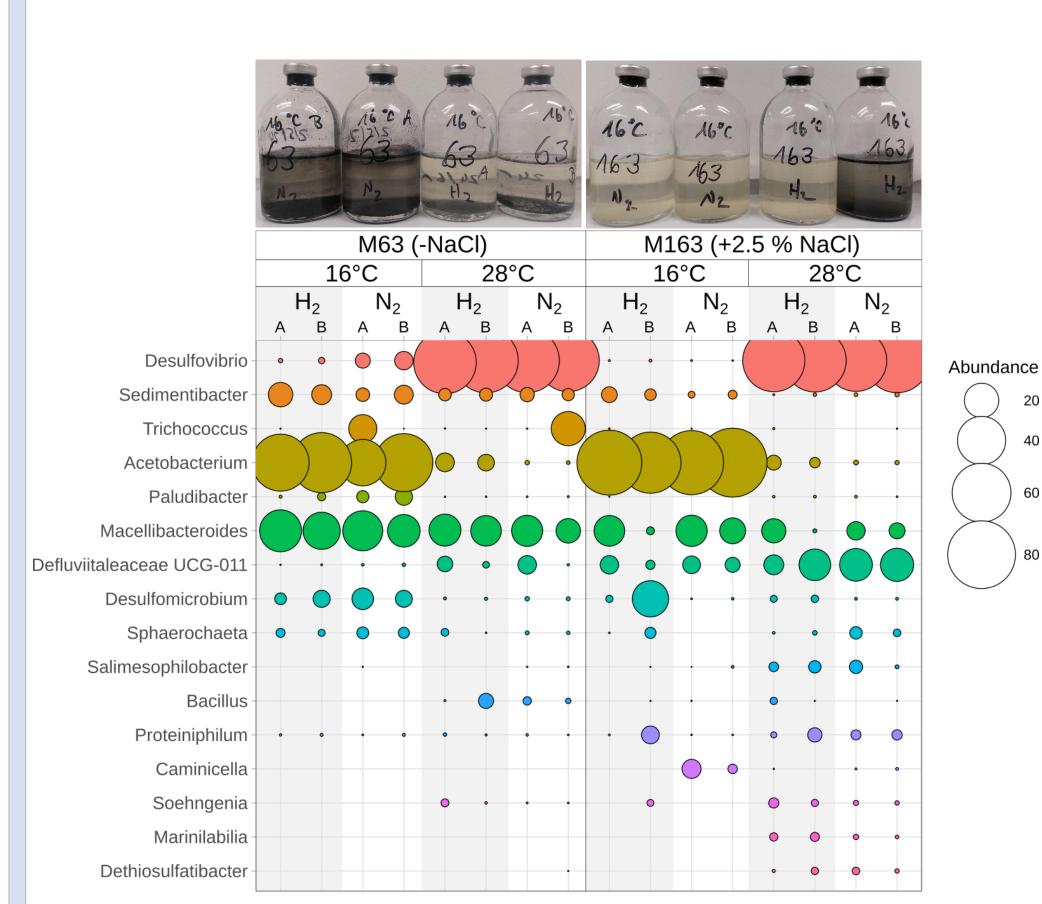


Fig. 3: Composition and relative abundance of taxa after enrichment for sulfate reducers at different temperatures and head spaces with and without NaCl.

- Abundance of SRB correlates with formation of black precipitates (most likely Fe-sulfide)
- Temperature controls the type of organisms being enriched
- No enrichment at 55 °C
- Salinity and head space composition have no significant impact